

# D-Fructose/D-Glucose, UV method

**Alternative Procedures** 

## **Total reducing sugars**

This kit allows the quantification of D-fructose and D-glucose individually. However, in many cases it is unnecessary to differentiate between these sugars, allowing them to be quantified together (total reducing sugars) using a more rapid assay format, as follows:

#### Supplementary preparation step:

Gently shake Suspensions 3 and 4 to remove any enzyme that may have settled on the caps. Using a pipette, transfer the entire contents of Suspension 4 (PGI) into Suspension 3 (HK/G6P-DH). Mix the enzymes by gentle swirling. This HK/ G6P-DH/PGI mixture is now ready for use.

After performing this step, D-glucose and D-fructose cannot be measured individually with this kit reagent mixture.

#### Procedure

Wavelength: 340 nm

Cuvette: 1 cm light path (glass or plastic)

Temperature: ~ 25 °C

Final volume: 2.34 mL (D-glucose plus D-fructose) Sample solution: 4-80  $\mu g$  of D-glucose plus D-fructose per cuvette (in 0.10-2.00 mL sample volume)

Read against air (without a cuvette in the light path) or against water

Pipette into cuvettes (mL)	Blank	Sample	
Distilled water (at ~25 °C)	2.10	2.00	
Sample	-	0.10	
Solution 1 (imidazole buffer)	0.10	0.10	
Solution 2 (NADP++ATP)	0.10	0.10	
Mix, measure the absorbance of the above solutions (A1) after approx. 3 min and start the reaction by addition of			
Suspension 3+4	0.04	0.04	
(HK+G6PDH + PGI))			
Mix* measure the absorbance of the solutions $(\Lambda,)$ at the			

Mix\*, measure the absorbance of the solutions (A<sub>total</sub>) at the end of the reaction (approx. 10 min). If the reaction has not stopped after 10 min, continue to read the absorbance at 2 min intervals until the absorbance remain the same over 2 min\*\*

### Calculation

Determine the absorbances difference for both blank and sample (A<sub>total</sub>-A<sub>1</sub>). Subtract the absorbances difference of the blank from the absorbance difference of the sample, thereby obtaining  $\Delta A_{D.}$  fructose +D-glucose. The concentration of Total reducing sugars (D-fructose+D-glucose; g/L), based on the Abs of NADH at 340 nm (6300 L×mol<sup>-1</sup>×cm<sup>-1</sup>), are calculated as follows:

# $C = 0.6692 \times \Delta A_{D-fructose + D-Glucose}$ [g/L]

# **Micro-volumes formats**

This kit has been developed to work in cuvettes with a standard pathlength of 1 cm, as described in the respective "Product Brochure". However, it can be adapted for use in 96-well microplates or in auto-analysers (micro-volume formats) with minimal assay optimisation. Basically, the assay volumes for the cuvette format have to be reduced approximately 10-fold for use in microplate format or in auto-analyser format. However, when using these micro-volume formats, you must be aware that the radiation pathlength is usually smaller than the standard cuvette pathlength of 1 cm. Thus, to perform the calculation of the amount of analyte in the samples under analysis follow one of the three strategies described in the section below.

#### Auto-analyser procedure (for reducing sugars)

This kit is appropriate for the preparation of 254.1 mL of reagent (equivalent to 1155 reactions of 0.230 mL). Reagent preparation is accomplished as follows:

Preparation of R1:

Component	Volume
Solution 1	1.0 mL
Solution 2 (after addition of 12 mL of H2O) Suspension 4 (swirl before use)	1.0 mL
	0.2 mL
	1.0 mL
PVP solution (10 mg/mL) or Distilled water	18.0 mL
Distilled water	

#### Preparation of R2:

Component	Volume
Suspension 3 (swirl before use)	0.2 mL
Distilled water	1.9 mL
T	otal 2.1 mL

**Example Procedure:** 

	Volume
R1	0.200 mL
Sample	0.002 mL*
R2	0.020 mL

Reaction time: 10 min at 25 °C or 5 min at 37 °C

Wavelength: 340 nm

**Prepared reagent stability:** > 7 days when refrigerated

Calculation: endpoint

Reaction direction: increase

**Linearity:** up to 108  $\mu\text{g}/\text{mL}$  of D-glucose + D-fructose in final reaction mixture

 $\ast$  If AU values are higher than 2, please dilute the sample with distilled water accordingly.

#### Strategies for analyte calculation

Auto-analysers use reaction volumes of approximately 0.315 mL and pathlengths from 4 to 8 mm, which is similar to a standard 96well microplate in which the same reaction volume would have a pathlength of 6 or 7 mm (similar assay volumes). Therefore, in both formats (96-well microplate and auto-analysers systems), the calculation of the analyte must be done by one of the three possible methods described below:

#### 1. Using the pathlength conversion factor

This is the easiest method to perform the calculation of the analyte. However, it requires a microplate reader with pathlength conversion capacity, i.e., the apparatus can detect the pathlength of each well and convert the individual readings to a 1 cm pathlength (cuvette format). In the case of auto-analysers, the absorbance readings should be directly converted to a 1 cm pathlength. This will allow the calculation of the analyte content as described in the "Product Brochure", provided with the kit and available at the NZYTech website.

#### 2. Using one standard curve

In this method, it is necessary to perform a standard curve of the analyte on each microplate that contains the test samples, or in the auto-analyser, and calculate the result from the standard curve of analyte concentration vs. absorbance. The standard curve can be performed by using the control solution provided in the kit.

#### 3. Using two standard curves

The most complicated method is to perform standard curves of the analyte in both the cuvette format (i.e. with a 1 cm of radiation pathlength) and the 96-well microplate or auto-analyser formats, and use these results to obtain a mean conversion factor between the cuvette procedure values and the alternative procedure values. The standard curves can be performed by using the control solution provided in the kit.

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