

# NZY First-Strand cDNA Synthesis Kit

## Catalogue number:

MB12501, 50 reactions

MB12502, 250 reactions

### Features

- Provides high yields of full-length cDNA products for use in RT-qPCR and two-step RT-PCR assays
- Formulated to increase sensitivity in RT-qPCR
- Primer type: oligo(dT)<sub>18</sub> and random hexamers
- Starting material: 10 pg to 5 µg of total RNA
- Optimal reaction temperature: 50 °C
- Convenient and reliable

### Description

NZY First-Strand cDNA Synthesis Kit is a system that includes all the necessary components to synthesize first-strand cDNA, except the template RNA.

The resulting single-stranded cDNA is suitable for use in real-time quantitative Reverse Transcription PCR (RT-qPCR). NZY First-Strand cDNA Synthesis Kit is formulated to provide high yields of full-length cDNA products and to increase sensitivity in RT-qPCR.

Starting material can range from 10 pg up to 5 µg of total RNA. The kit includes a combination of random hexamers and oligo(dT)<sub>18</sub> primers in order to increase sensitivity. The primers are included in the NZYRT 2× Master Mix, which also contains dNTPs, MgCl<sub>2</sub> and an optimized RT buffer. NZYRT Enzyme Mix includes both NZY Reverse Transcriptase (RNase H minus) and NZY Ribonuclease Inhibitor in order to protect RNA against degradation due to ribonuclease contamination. RNase H (from *E. coli*) is provided in a separate tube to specifically degrade the RNA template in cDNA-RNA hybrids after the first-strand cDNA synthesis. This procedure will improve the sensitivity of subsequent RT-qPCR reaction since PCR primers will bind more easily to the cDNA.

### Storage conditions

Store all kit components at -20 °C in a freezer without defrost cycles. Stability can be extended by storing it at -80 °C. The kit will remain stable till the expiry date if stored as specified.

## System Components

Component	MB12501 (50 reactions)	MB12502 (250 reactions)
NZYRT Enzyme Mix <sup>(1)</sup>	100 µL	5 × 100 µL
NZYRT 2× Master Mix <sup>(2)</sup>	500 µL	5 × 500 µL
NZY RNase H ( <i>E. coli</i> )	50 µL	5 × 50 µL
DEPC-treated H <sub>2</sub> O	1 mL	2 × 1 mL

(1) Includes NZY Reverse Transcriptase and NZY Ribonuclease Inhibitor

(2) Includes oligo(dT)<sub>18</sub>, random hexamers, MgCl<sub>2</sub> and dNTPs

## Protocol for first-strand cDNA synthesis

1. On ice, add the following reaction components into a sterile, nuclease-free microcentrifuge tube (for multiple reactions, a master mix without RNA may be prepared):

NZYRT 2× Master Mix	10 µL
NZYRT Enzyme Mix	2 µL
RNA (up to 5 µg)	× µL
DEPC-treated H <sub>2</sub> O	up to 20 µL

2. Mix gently and incubate at 25 °C for 10 min.

3. Incubate at 50 °C for 30 min.

4. Inactivate the reaction by heating at 85 °C for 5 min, and then chill on ice.

5. Add 1 µL of NZY RNase H and incubate at 37 °C for 20 min.

**Note:** Addition of NZY RNase H will remove RNA bond to cDNA. This procedure is mainly recommended when using cDNA in PCR amplification, especially for some targets (> 1 kb) that may require RNA-free DNA as template. RNA removal will increase the sensitivity of the PCR step.

6. Use the cDNA product directly in PCR or qPCR diluted in TE buffer or undiluted; or store at -20 °C until required.

## Important notes

- High quality intact RNA, free of residual genomic DNA and RNases is essential for full-length, high quality cDNA synthesis and accurate RNA quantification. For this reason, special precautions should be taken when working with RNA:
  - Aseptic conditions should be maintained: always wear gloves; change gloves whenever you suspect that they are contaminated; use RNase-free tubes and pipet tips; designate a special area and equipment for RNA work only.
  - DNase I (not provided) may be used to eliminate genomic DNA contamination from the starting total RNA.
  - The template RNA should be stored at -70 °C. Avoid multiple freeze/thaw cycles of RNA.
- This kit does not include control RNA.
- Keep all reagents of the kit on ice while setting up the reactions.
- When performing RT-qPCR using the synthesized cDNA as template, no more than 1/10 of the final PCR volume should be derived from the reverse-transcription product. For instance, use up to 5 µL of cDNA obtained in the first-strand synthesis in a 50 µL PCR reaction.

## Quality control assays

### Purity

NZY Reverse Transcriptase and NZY Ribonuclease Inhibitor present in the NZYRT Enzyme Mix are >90% pure as judged by SDS polyacrylamide gel electrophoresis followed by BlueSafe staining.

### Nucleases assays

All components of the kit are tested for DNase and RNase contamination, using 0.2-0.3 µg of pNZY28 plasmid DNA and 1 µg of RNA, respectively. Following incubation at 37 °C, the nucleic acids are visualized on a GreenSafe-stained agarose gel. There must be no visible nicking or cutting of the nucleic acids.

### Functional assay

NZY First-Strand cDNA Synthesis Kit is tested for performance in a RT-qPCR experiment using a 10-fold serial dilution of total RNA from mouse liver (1 µg to 0.1 ng). The resultant cDNA is then used as template in a quantitative real-time PCR assay using specific primers to amplify the mouse GAPDH gene.

## Troubleshooting

### Little or no RT-PCR/RT-qPCR amplification product

- RNA damage or degradation

Analyse RNA on a denaturing gel to verify integrity. Use aseptic conditions while working with RNA to prevent RNase contamination. Replace RNA if necessary.

- Presence of RT inhibitors

Some inhibitors of RT enzymes include: SDS, EDTA, glycerol, sodium phosphate, spermidine, formamide and guanidine salts. Remove inhibitors by ethanol precipitation of the RNA preparation before use; wash the pellet with 70% (v/v) ethanol.

- Not enough starting RNA

Increase the concentration of starting RNA.

### Unexpected bands after electrophoretic analysis of amplified products

- Genomic DNA contamination

DNase I may be used to eliminate genomic DNA contamination from the starting RNA.

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## Certificate of Analysis

Test	Result
Enzyme purity	Pass
Nucleases assays	Pass
Functional assay	Pass

Approved by:



Patrícia Ponte  
Senior Manager, Quality Systems

For research use only.

