MB135_SUP5_EN_V2401

NZY Tissue gDNA Isolation Kit

Catalogue number	Presentation
MB13502	50 columns

Support Protocol for the isolation of genomic DNA from Buccal Swabs

1. Sample preparation

Collect the samples with cotton, dracon® (Daigger), or C.E.P swabs (Gibco BRL). Scrape firmly against the inside of each cheek several times and let the swabs air dry.

2. Sample Pre-lysis

Place the dry swab material in 2 mL microcentrifuge tubes. Add 400-600 µL PBS buffer (not provided) and 25 µL Proteinase K solution to the swabs. Mix by vortexing 2 x 10 s and incubate at 56 °C for 10 min.

Note: *The volume of PBS buffer is dependent on the type of swab used: for cotton and dacron® swabs, 400 µL are sufficient; for C.E.P. swabs: 600 µL are necessary.*

Transfer as much as possible of the lysate solution to a 2 mL microcentrifuge tube (not provided). Discard swab and continue with recovered solution.

3. Sample Lysis

Add one volume Buffer NL (400 µL or 600 µL – depending on the swab type/volume of PBS buffer used) and vortex vigorously. Incubate the samples at 70 °C for 10 min.

4. Addition of Ethanol

Add one volume 96-100% ethanol (400 µL or 600 µL – depending on the swab type) to each sample and mix by vortexing.

5. DNA Binding

Transfer 600 µL of the samples from the 2 ml microcentrifuge into NZYSpin Tissue columns. Centrifuge at 11,000 x g for 1 min. If the samples are not drawn through completely, repeat the centrifugation. Discard flow-through.

Place the columns back into the collection tubes and repeat step 5, depending on the lysis volume.

Proceed with step 7 of the standard protocol.

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