

NZYBradford reagent

Catalogue number: MB19801, 500 mL

Description

The NZYBradford reagent for the determination of total protein is based on the property of Brilliant Blue G-250 to bind to proteins. When this dye binds to the basic and aromatic aminoacids of proteins, its absorbance maximum is shifted from 465 nm to 595 nm. This absorbance at 595 nm is proportional to the protein concentration.

Storage temperature

Each bottle contains 500 mL of the ready-to-use reagent solution and is sufficient for approximately 200 determinations, as stated in the protocol. The solution is stable for at least 2 years if kept between + 2 and + 8 °C in a refrigerator. At room temperature, it is stable for about 12 months.

Measuring ranges

Range 1: 0.1 – 1.4 mg/mL

Range 2: 0.01 – 0.1 mg/mL

Sample preparation

Turbid samples should be centrifuged or filtered. If the protein concentration is high, the sample should be diluted so that measurements can be carried out within the measuring range. Use the table below as a guide:

Suspected protein concentration (mg/mL)	
0.01 – 0.1	Measuring range 2
0.1 – 1.4	Measuring range 1
>1.4	dilution

Preparation of standard solutions

Bovine serum albumin (BSA) is frequently used as a reference protein. Prepare a BSA standard solution by dissolving precisely 100 mg of BSA (MB046) in a final volume of 10 mL of redistilled water. This stock solution (10 mg/mL) can be diluted as required:

Measuring range 1 (0.1 – 1.4 mg/mL)		
Standard solutions (mg/mL)	Protein stock solution (10 mg/mL)	Redistilled water (mL)
0.2	0.20	9.80
0.4	0.40	9.60
0.6	0.60	9.40
0.8	0.80	9.20
1.0	1.00	9.00
1.2	1.20	8.80
1.4	1.40	8.60

Measuring range 2 (0.01 – 0.1 mg/mL)		
Standard solutions (mg/mL)	Protein stock solution II (0.1 mg/mL)	Redistilled water (mL)
0.01	0.1	0.9
0.02	0.2	0.8
0.04	0.4	0.6
0.06	0.6	0.4
0.08	0.8	0.2
0.10	1.0	-

Protein stock solution II: Place 0.1 mL of the 10.0 mg/mL standard solution in a 10 mL volumetric flask and make up to the mark with redistilled water.

Standard Protocol

Measurements should be carried out in a spectrophotometer using a disposable plastic or glass cell (path length 1 cm) at 595 nm. Quartz cells are not advised since the dye adsorbs strongly onto this surface. Zero adjustment of the photometer can be carried out against air or water

Pipetting scheme for cell test Measuring range 1 (0.1 – 1.4 mg/mL)	Sample or standard	Reagent blank
Sample solution /standard solution	0.05 mL	-
Redistilled water	-	0.05 mL
Bradford reagent (solution)	2.5 mL	2.5 mL
Mix thoroughly, wait 2 minutes and measure absorbance at 595 nm.		

Pipetting scheme for cell test Measuring range 2 (0.01 – 0.1 mg/mL)	Sample or standard	Reagent blank
Sample solution /standard solution	0.25 mL	-
Redistilled water	-	0.25 mL
Bradford reagent (solution)	2.5 mL	2.5 mL
Mix thoroughly, wait 2 minutes and measure absorbance at 595 nm.		

The test can also be performed using microtitre plates, although the volumes and measuring range have to be changed. Also, precision and reproducibility have been shown to be worse than when using a conventional cell test due to the relatively high pipetting error involved.

Pipetting scheme for microtitre plates Measuring range 1 (0.05 – 0.5 mg/mL)	Sample or standard	Reagent blank
Sample solution /standard solution	10 µL	-
Redistilled water	-	10 µL
Bradford reagent (solution)	200 µL	200 µL
Mix thoroughly, wait 2 minutes and measure absorbance at 595 nm.		

Calculation

To compile a calibration curve, subtract the absorbance of the reagent blank from that of the standard. The difference ΔE can then be plotted against the standard protein concentrations.

$$\Delta E_{\text{Standard}} = E_{\text{Standard}} - E_{\text{Blank}}$$

or

$$\Delta E_{\text{Sample}} = E_{\text{Sample}} - E_{\text{Blank}}$$

The calibration curve is rarely linear over the entire concentration range. Thus, the protein concentration of an unknown sample should be calculated either graphically or by means of linear regression.

NB: If the sample has been diluted prior to measurement, the result must be multiplied by the appropriate dilution factor f .

Interference

Bradford Protein assay readings may be affected by some reagents present in the sample. The table below shows which reagents and what concentrations can cause interference. Combinations of these reagents can give rise to other reactions that also interfere with the test. To avoid interferences it is recommended preparing all blank and standard solutions with the same buffer used in the preparation of the sample.

Inorganic salts	
Ammonium sulphate	>1 M
KCl	>1 M
MgCl ₂	>1 M
Na azide	>0.5%
NaCl	>5 M
NaSCN	>3 M
Detergents	
Brij	>0.5%
Desoxycholate	>0.1%
SDS	>0.1%
Triton X-100	>0.1%
Tween 20	>0.5%
Nucleic acids, etc	
Adenosine	>1 mM
ATP	>1 mM
DNA	>1 mg/mL
RNA	>0.3 mg/mL
rRNA	>0.25 mg/mL
tRNA	>0.4 mg/mL
Thymidine	>1 mM

Buffers	
Acetate	>0.5 M
BES	>2.5 M
CHAPS	>1%
CHAPSO	>1%
Citrate	>50 mM
Glycine	>0.1 M
HEPES	>0.1 M
MES	>0.7 M
MOPS	>0.2 M
Phosphate	>1 M
PIPES	>0.5 M
Resolytes	>0.5%
Tris	>2 M

V1902

Certificate of Analysis

Assay	Result
Protein Standard Curve (BSA, 595 nm)	Pass

Approved by:



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Senior Manager, Quality Systems

For research use only.

