

NZYProof 2× Colourless Master Mix

Catalogue number: MB28801, 500 U
 MB28802, 1000 U
 MB28803, 5000 U

Description

NZYProof 2× Colourless Master Mix is a premixed ready-to-use solution containing NZYProof DNA polymerase, an enzyme that presents high fidelity with robust activity and displays great performance in most PCR applications. NZYProof DNA polymerase possesses 3'→5' exonuclease proofreading capacity which enables the polymerase to amplify DNA with increased accuracy. The enzyme is highly efficient in the amplification of longer (≤10 kb) PCR products and site-directed mutagenesis. In addition, it is the recommended polymerase for routine cloning that requires precision. The master mix was optimized to allow high yields and enhanced sensitivity due to the presence of stabilizer solution. It contains dNTPs, reaction buffer and additives at optimal concentrations for the accurate amplification of a large range of DNA templates by PCR. Mg²⁺ final concentration is 1.5 mM, allowing the implementation of a variety of PCR protocols. For highly sensitive downstream applications it is recommended to purify the PCR products using NZYGelpure (MB011) before usage in subsequent protocols. NZYProof DNA polymerase possesses 3'→5' exonuclease proofreading activity. Resulting PCR products have blunt-ends and are suitable for cloning with Nzytech's NZY-blunt PCR cloning kit (MB121).

Shipping conditions

The product can be shipped in a range of temperatures from dry ice to blue ice.

Storage conditions

This master mix should be stored at -85 °C to -15 °C in a freezer without defrost cycles to guarantee maximal shelf life. Minimize the number of freeze-thaw cycles by storing in working aliquots. The product will remain stable till the expiry date if stored as specified.

Unit definition

One unit is defined as the amount of enzyme required to catalyse the incorporation of 10 nmoles of dNTPs into acid insoluble material in 30 minutes at 72 °C, under the following assay conditions: 50 mM Tris-HCl, pH 9.0, 50 mM NaCl, 2.5 mM MgCl₂, 200 μM each of dATP, dCTP, dGTP, dTTP (a mix of unlabelled and α-[³²P]-labelled), 10 μg activated salmon sperm DNA in a final volume of 50 μL.

Enzyme concentration: 0.2 U/μL

Standard Protocol

The following standard protocol serves as a general guideline and a starting point for any PCR amplification. Optimal reaction conditions (incubation times and temperatures, concentration of primers and template DNA) may vary, although PCR optimization is usually not required. In case you need to fine-tune primer concentrations, test the recommended variations provided in brackets in the table below.

1. Gently mix and briefly centrifuge the master mix after thawing. Set up the PCR reaction on ice and add water first and the remaining components in the order specified in the table below. A single reaction mixture of 50 μL should combine the following components:

Forward and Reverse Primers	0.4 (0.3-0.5) μM
Template DNA	5 ng-0.5 μg
NZYProof 2× Colourless Master Mix,	25 μL
Nuclease-free water	up to 50 μL

2. Mix and quickly pulse the reactions.

3. Immediately initiate the PCR by transferring the PCR mixtures to the thermocycler with the block pre-heated to 95 °C and following the below cycling parameters:

Cycle step	Temp.	Time	Cycles
Initial denaturation	95 °C	3 min	1
Denaturation	95 °C	30 s	20-40
Annealing	*	30 s	
Extension	72 °C	60 s/kb	
Final Extension	72 °C	5-10 min	1

*Annealing temperature should be optimized for each primer set based on the primer T_m; typically it should be T_m -5 °C.

4. Analyse PCR products by agarose gel electrophoresis (0.7-1.2%, w/v) and visualize with GreenSafe Premium (MB132) or any other mean.

Primer Design

PCR primers generally range in length from 15–30 bases and are designed to flank the region of interest. Sequences longer than 30bp may improve PCR yield using NZYProof DNA polymerase since its 3'→5' exonuclease activity may degrade primers. In addition, to overcome primer degradation, the 3' termini of primers may be protected with phosphorothioate modifications. Primers should contain 40–60% GC and avoid sequences that might produce internal secondary structure. The 3'-ends of the primers should not be complementary to avoid the production of primer-dimers. Primer-dimers unnecessarily remove primers from the reaction and result in an unwanted polymerase reaction that competes with the desired reaction. Avoid three G or C nucleotides in a row near the 3'-end of the primer as this may result in non-specific primer annealing increasing the synthesis of undesirable reaction products. Ideally, both primers should have nearly identical melting temperatures (T_m); in this manner, the two primers will anneal at roughly the same temperature.

DNA template

The optimal amount of starting material may vary depending on its quality and complexity. In general, we recommend using 50 ng to 500 ng of genomic DNA templates, although the enzyme is sensitive

enough to amplify fragments from as little as 5 ng of human gDNA, for example. Lower amounts of template may be used for amplification of less complex DNA (typically 10-50 ng). When using a cDNA synthesis reaction as template do not exceed 10% of the final PCR reaction volume.

Quality control assays

Purity

NZYProof DNA polymerase purity is > 90% as judged by SDS-PAGE followed by Coomassie Blue staining.

Genomic DNA contamination

The product must be free of any detectable DNA contamination as evaluated through PCR. Thus, it is suitable for the amplification of bacterial and fungal DNA based on 16S and 18S rRNA PCR assays.

Nuclease assays

0.2-0.3 µg of pNZY28 plasmid DNA are incubated with NZYProof 2× Colourless Master Mix for 14-16 hours at 37 °C. Following incubation, the DNA is visualised on a GreenSafe Premium-stained agarose gel. There must be no visible nicking or cutting of the nucleic acid.

Functional assay

NZYProof 2× Colourless Master Mix is tested for performance in a PCR reaction using 1.25 units of enzyme for the amplification of different-sized DNA fragments (1 and 2.5 kb) from human genomic DNA. The resulting PCR products are visualized as a single band in a GreenSafe stained agarose gel.

Troubleshooting

No product amplification or low yield

- Inadequate annealing temperature

The reaction mix composition may affect the melting properties of primers and DNA. Adjust the annealing temperature to accommodate the primer with the lowest melting temperature (5 ° to 10 °C lower than T_m).

- Presence of PCR inhibitors

Some DNA isolation procedures, particularly genomic DNA isolation, can result in the co-purification of PCR inhibitors. Reduce the volume of template DNA in reaction or dilute template DNA prior to adding to the reaction. Diluting samples even 1:10,000 has been shown to be effective in improving results, depending on initial DNA concentration.

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