

LyoNZYSupremeMultiplex One-Step RT-qPCR Probe Master Mix (2)

Catalogue number:

MB44301, for 1.5 mL
(150 x 20 µL rxs)

Description

LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix is an optimized and highly efficient freeze-dried reaction mixture developed for first-strand cDNA synthesis and subsequent real-time PCR in a single tube. This offers great convenience and minimizes the risk of errors and contaminations. The master mix, formulated as a 2× reaction mixture, contains all components necessary for both cDNA synthesis and multiplex qPCR amplification (including enzymes, dNTPs, stabilizers and enhancers). Depending on the real-time thermal cycler used, up to 5-6 targets can be quantified simultaneously in the same well or tube. This master mix was engineered with a dual hot-start enzyme control mechanism to provide the highest detection sensitivity. In addition, the latest developments in PCR enhancers have been incorporated in the LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix, including buffer chemistry and incorporation of highly robust engineered enzymes mixed in an optimized proportion specifically designed for the highest-performance. This master mix does not contain ROX and it was specifically developed for probe-detection technology. For qPCR instruments that require this reference dye, we indicate a concentration range for your assay according to the table shown in the section "ROX reference dye" (ROX not provided).

Shipping Conditions

The product can be shipped in a range of temperatures from room temperature to dry ice.

Storage Conditions

Although the product components provided are stable at room temperature, upon arrival they should be stored at -85 °C to -15 °C in a freezer without defrost cycles in order to guarantee maximal shelf life. Short-term storage for 1-2 weeks at room temperature will not compromise the performance of the product. Once the lyophilized master mix has been resuspended using the buffer provided, it should be stored for short-term at -85 °C to -15 °C. Minimize the number of freeze-thaw cycles by storing it in working aliquots. The master mix will remain stable till the expiry date if stored as specified.

Pack components

Component	MB41701
LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix (2x)	3 tubes with lyophilized master mix
Multiplex RT-qPCR master mix reconstitution buffer	1.75 mL

Preparation of LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix

Reconstitute each tube of LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix with 525 µL of the Multiplex RT-qPCR master mix reconstitution buffer provided. Complete hydration of the mixture might take 2-3 min. Mix gently by flicking the tube, and spin down. Do not replace the reconstitution buffer with water or any other buffer. The master mix is then ready to use as a 2x qPCR master mix.

Compatible real-time PCR instruments

The master mix is compatible with real-time PCR instruments that do not require a passive reference signal for data normalization. It has been optimized to be compatible with the following real-time PCR instruments:

Bio-Rad®:

CFX96™; CFX384™; iCycler®; iQ™5; Opticon™; Opticon™ 2

Qiagen (Corbett):

Rotor-Gene™ 3000; Rotor-Gene™ 6000 & Rotor-Gene™ Q

Roche:

Lightcycler® 96; Lightcycler® 480 & Lightcycler® Nano

Applied Biosystems (with optional ROX addition):

7000; 7300; 7700; 7900; 7900HT; 7900HT FAST; StepOne™ & StepOne™plus; 7500; 7500 FAST; QuantStudio™ 5, 6, 7, 12k Flex & ViiA7™

Protocol

The following protocol serves as a general guideline and a starting point for any One-step RT-qPCR procedure. Optimal reaction conditions (e.g., incubation times, temperatures, and primer/template concentration) may vary and, in particular conditions, may require further optimization.

RT-qPCR reaction set-up: the given volumes are based on a standard 20 µL final reaction mix and can be scale adjusted.

LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix (2x) ^(*) ^(*)	10 µL	1×
Each forward primer (at 10 µM)	0.8 µL	400 nM ^(*)
Each reverse primer (at 10 µM)	0.8 µL	400 nM ^(*)
Each probe (at 10 µM)	0.4 µL	200 nM ^(*)
RNA Template	up to 8 µL	-
Nuclease-free water	up to 20 µL	-

(Alternative set-up) For easy handling of multiple primers used in multiplex assays, we recommend preparing primer/probe mixes for each target by combining the different sets of primers and probes at required concentrations. For example, for a double gene targeting multiplex qPCR, we recommend preparing two 10x concentrated mixes of primers and probes for both targets and proceed as following (*note: if it is more convenient, alternatively*

prepare 20x concentrated mixes, and then use 1 µL in the reaction mix):

LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix (2x) ^(*) ^(**)	10 µL	1x
10x Primer + probe mix - target 1 (see Table 1)	2 µL	primers: 400 nM ^(*) ; probe: 200 nM ^(*)
10x Primer + probe mix - target 2 (see Table 1)	2 µL	primers: 400 nM ^(*) ; probe: 200 nM ^(*)
RNA Template	up to 6 µL	-
Nuclease-free water	up to 20 µL	-

Table 1: Concentrated assay mixtures of primers and probes with suggested concentrations for a standard qPCR reaction ^(*)

Mix (final)	Primer Forward	Primer Reverse	Probe
10x	4 µM	4 µM	2 µM
20x	8 µM	8 µM	4 µM

^(*) The master mix does not contain ROX, but, if required, addition of this internal passive reference dye can be conducted in a separate step. The final concentration will vary according to the qPCR instrument used. Please follow instructions described in the section "ROX reference dye" below.

^(**) Please notice that a precipitate in the bottom of the master mix tube may be observed, in particular after multiple freeze/thaw cycles. To ensure optimal performance, please make sure all components are thawed and resuspended/homogenized prior to use. In this case do not spin down the master mix before pipetting.

^(*) See section of "General considerations" below for more details about primers and probes final concentrations in the reaction.

Testing and Ct values: When comparing this RT-qPCR master mix with a mix from another supplier we strongly recommend amplifying from a 10-fold template dilution series. Loss of detection at low template concentration is the only direct measurement of sensitivity. An early Ct value is not an indication of good sensitivity, but rather an indication of speed.

ROX reference dye: LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix (2x) is compatible with most thermocyclers available in the market and can include ROX passive reference dye (not provided) to normalize non-PCR-related fluctuations in fluorescence. If ROX addition is required for your qPCR platform, an optimal quantity of this dye should be included in your master mix. The recommended amount of ROX for the most common qPCR instruments is stated in the table below:

qPCR instrument	Volume of ROX per 1 mL of Master Mix (2x) ^(*)
Applied Biosystems: 7000/7300/7700/7900/7900HT/7900HT FAST/StepOne™/StepOne™plus	20-29 µL (500-725 nM)
Applied Biosystems: 7500/7500FAST/QuantStudio™ 5, 6, 7, 12k Flex/ViiA7™	2-4 µL (50-100 nM)
Bio-Rad®: CFX96™/CFX384™/iCycler®/iQ™5/Opticon™/Opticon™ 2 Qiagen: Rotor-Gene™ 3000/6000/Q Roche: Lightcycler® 96/480/Nano	Not required

^(*) For different volumes please scale-up or scale-down the volume of ROX accordingly. Please notice that ROX is not provided by NZYtech.

Suggested thermal cycling conditions

LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix was optimized for the amplification of RNA fragments up to 200 bp under different RT-qPCR cycling conditions. The table below displays a standard cycling setup optimized on several platforms. However, these conditions may be adapted to suit different machine-specific protocols.

Cycles	Temp.	Time	Main reaction
1	50 °C	10-20 min ^(*)	Reverse Transcription
1	95 °C	2-5 min	Polymerase activation
40	95 °C	5 sec	Denaturation
	60 °C	30 sec-50 sec	Annealing/Extension

^(*) Reverse transcriptase has a high processivity and may take as little as 10 minutes to synthesize cDNA. However, in specific situations increasing reaction time up to 20 minutes may be beneficial.

General considerations

The inherent chemical instability of RNA and the ubiquitous presence of RNases require that particular care should be taken while working with RNA. We recommend using RNase-free plasticware/reagents and work in an RNase-free area (RNase Cleaner, Cat. No. MB16001, can help removing RNases from surfaces and materials). In addition, to help prevent any carry-over DNA contamination, you should assign independent areas for reaction set-up, PCR amplification and any post-PCR gel analysis. It is essential that any tubes containing amplified PCR product are not opened in the PCR set-up area.

Primers and probe: These guidelines refer to the design and set-up of dual labelled probes. Please refer to the relevant literature when using other probe types. The specific amplification, yield and overall efficiency of any real-time RT-PCR can be critically affected by the sequence and concentration of the probes and primers, as well as by the amplicon length. We strongly recommend taking the following points into consideration when designing and running your real-time RT-qPCR experiment:

- Primers should have a melting temperature (T_m) of approximately 60 °C. The probe T_m should be approximately 10°C higher than that of the primers;
- **Use spectrally distinct fluorophores to label each probe;**
- The fragment to amplify should be between 80-200 bp in length and not superior to 300 bp;
- Final primer concentrations of 400 nM are suitable for most probe-based reactions. However, to determine the optimal concentration we recommend titrating in the range 0.2-1 µM. Forward and reverse primers concentration should be equimolar;
- A final probe concentration of 100 nM is suitable for most applications; we recommend that the final probe concentration is at least two-fold lower than the primer concentration; to determine the optimal concentration we recommend titrating in the range 0.1-0.25 µM;
- For multiplex RT-qPCR, probe concentrations higher than 100 nM can result in cross channel fluorescence.
- Prior to any multiplex assay, single qPCR reactions might be performed to test the functionality and specificity of each primer/probe pair.

Template: It is important that the RNA template is purified and devoid of contamination by RT-qPCR inhibitors (e.g., EDTA). The recommended amount of template is dependent upon the type of RNA used. Please consider the following points when selecting RNA templates:

- **Total RNA:** purified total RNA can be used in the range from 1 pg to 1 µg per 20 µL reaction;
- **mRNA:** purified mRNA can be used from 0.01 pg per 20 µL reaction.

To obtain high yield of highly purified RNA we suggest using the NZY Viral RNA Isolation kit (Cat. No. MB407) or NZY Total RNA Isolation Kit (Cat. No. MB134).

Mg²⁺: It is not necessary to supplement the reaction mixture with MgCl₂ as the LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix already contains an optimized concentration of MgCl₂.

RT-qPCR optimization: It may be necessary to improve the efficiency of some reactions, such as multiplexing with more than two probes, or if the target amplicon is longer than 200 bp. In these cases, the reverse transcription reaction time can be extended up to 30 minutes; the annealing/extension time can be extended up to 60 seconds.

Quality control assays

Genomic DNA contamination

The product must comply with internal standards of DNA contamination as evaluated through real-time qPCR.

Nuclease assays

To test for DNase contamination, 0.2-0.3 µg of pNZY28 plasmid DNA are incubated with the master mix for 14-16 h at 37 °C. To test for RNase contamination, 1 µg of RNA is incubated with the master mix for 1 h at 37 °C. Following incubation, the nucleic acids are visualized on a GreenSafe-stained agarose gel. There must be no visible nicking or cutting of the nucleic acids.

Functional assay

LyoNZYSupremeMultiplex One-step RT-qPCR Probe Master Mix is extensively tested for multiplexing activity, processivity, efficiency, sensitivity and heat activation.

V2401

For research use only.

For life science research only. Not for use in diagnostic procedures.